



Metabolic profile and biological activities of *Lavandula stoechas* L.

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Abstract: Medicinal and aromatic plants play a significant role in drug discovery. *Lavandula stoechas* L. has been used as folk medicine to treat various diseases. The aim of this work is to investigate the phytochemistry of *Lavandula stoechas* with biological activities. An aerial part of the plant was extracted with methanol. Another sample of plant was boiled in water then aqueous part was extracted with ethyl acetate. Identification and quantification of phenolic compounds, organic acids and flavonoids in methanol extract were carried out by High Performance Liquid Chromatography/Time of Flight/Mass Spectrometry, HPLC-TOF/MS. Rosmarinic acid was found as a chief compound (80.9%). The essential oil was generated by steam distillation and identified by GC-MS. The main constituents were camphor (48.1%) and fenchone (30.5%). The essential oil exhibited good insecticidal activity on *Sitophilus granarius* and *Sitophilus oryzae* pests as 43.3% and 62.9% mortality, respectively. However, the methanol extract has only insecticidal activity against *S. granaries* (50.0%). The antioxidant activities were investigated using assays of 1,1-diphenyl-2-picrylhydrazyl (DPPH[•]) radical scavenging, reducing power (FRAP), and 2,2'-azino-bis-(3-ethylbenzothiazoline-6-sulphonate) (ABTS^{•+}) radical scavenging on both extracts. In addition, total phenolic contents were determined. Total phenolic content of EtOAc extract was found as 2.18 g GAE (Gallic acid equivalent) phenolic compounds/kg plant. Whereas, total phenolic of methanol was 3.33 g GAE/kg plant. EtOAc extract revealed the considerable DPPH[•] scavenging, ABTS^{•+} scavenging and reducing power activities with the values of 28.71 (IC₅₀, µg/ml), 8.72 (IC₅₀, µg/ml) and 6.99 (µmol trolox equivalent/mg extract) respectively.

Key words: Chemical constituents; essential oil components; antioxidant activity; insecticidal activity; *Lavandula stoechas*.

Introduction

Natural products play a significant role in drug discovery and development process due to the including a large variety of biological active constituents (1-9).

The genus *Lavandula* (Family Lamiaceae) is considered a medicinal and aromatic plant with great economic values, cultivated in many European countries, widely distributed throughout the world and consists of 35 species (10). The *Lavandula* species have been utilized as an alternative medicine to treat diverse diseases such as diabetes, depression, headaches (11). The essential oils of *L. stoechas* are well documented and known to have anti-inflammatory, antifungal, antibacterial, antioxidant, sedative, neuroprotective activities. They have been used for food, cosmetic, perfumery and pharmaceutical industries as well as treatment of wounds, burns and skin injuries (12). The essential oils containing necrodane monoterpenoid inhibited beta secretase activity, so they exhibited the ability to penetrate cell membranes. Therefore, these oils have a potency to be a medicine for Alzheimer's disease (13). Besides essential oils, photochemical researches of *Lavandula* species have brought about the isolation of monoterpenes (14), flavonoids (15), dichotomosite, succinic acid, caffeic acid, ferulic acid, beta-sitosterol, ursolic acid and daucosterol (16). *L. stoechas* consists of acetylated glucoside of luteolin and flavone glucosides (17), triterpenes, steroids (18).

Essential oils (EOs) (volatile oils) are aromatic oily

liquids generated from plant materials. They are widely used in pharmaceutical and food industries. EOs have also been reported to display a wide range of biological activities. (19,20).

Insects have been known the most significant problem for agricultural industries. They damage to the crops while growing up and after harvest. Many works have been conducted to find the best solution to these issues. The synthetic chemicals used for insecticide can be harmful to the environment as well as human beings who consume chemically contaminated food (21). Natural products have attracted the great attention of scientists who search to find non-toxic, environmentally safe insecticide (22).

Sitophilus granaries (L.) (Coleoptera: Curculionidae) is one of the most dangerous pests effecting stored wheat kernels. Because of the invasion, nutritional wheat values deteriorate and an increase in hygiene problems can be observed (23).

Sitophilus oryzae (L.) (Coleoptera: Curculionidae) is considered to be one of the most destructive species in stored cereal grains throughout the world. It can attack intact grain. The adults feed on grains and the larva grows inside the grain kernels causing both quantitative and qualitative losses of the stored grain products (24).

Hydroxyl radicals, superoxide and singlet oxygen called the reactive oxygen species (ROS), responsible many chemical reactions in eukaryotic cells can harm cells leading to disease such as DNA damage, cardiovascular diseases and cancer (25). Antioxidants

have been used in food industry to keep the food freshly. The main source of deterioration, nutritional losses and discoloration in foods are free radicals oxidizing the antioxidant compounds. Recently, a great number of scientific investigations have been carried out to find the natural antioxidants for use in food industries to replace synthetic ones which have been restricted on account of their carcinogenicity (26-28).

Herein we extracted aerial part of *L. stoechas* with methanol and ethyl acetate. Chemical constituents of methanol extract were identified by LC-TOF/MS. The essential oil of *L. stoechas* was obtained by steam distillation and identified by GC-MS. Moreover, insecticidal activity of essential oil and methanol extract was carried out on *Sitophilus granarius* and *Sitophilus oryzae* pests and found out that the essential oil exhibited good insecticidal activity. Antioxidant activities of methanol and ethyl acetate extracts were carried out and these extracts exhibited the considerably antioxidant activities.

Materials and Methods

Chemical and Reagents

2,2'-Azino-bis(3-ethylbenzthiazoline-6-sulfonic acid) (ABTS), Butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), Trolox, 1,1-diphenyl-2-picryl-hydrazyl (DPPH[•]), trichloroacetic acid (TCA) were obtained from Sigma. Gallic acid, sodium carbonate, potassium ferricyanide, Folin-Ciocalteu reagent and solvents were bought from Merck.

Plant material

Lavandula stoechas L. was collected from Denizli, Turkey during the flowering season and identified by Prof. Dr. Ahmet Ilcim, Mustafa Kemal University, Faculty of Arts and Sciences, Department of Biology where a voucher specimen was deposited (No: MKUH-1329).

Insects

The cultures of *S. granarius* and *S. oryzae* were supplied from Cankiri Karatekin University, Biology Department. The wheat was filled into the one third of glassware (5.0 L), and then adult males and females were added to lay eggs to get a single aged population. The adults were removed and cultures were incubated at 27 °C and 60% relative humidity in a dark climate chamber after 48 h later. After forty-five days, the new generation of adults appeared to be used in experiments (22).

Extractions

The aerial part of plant material (100 g) was dried at room temperature at shadow. It was powdered then extracted with methanol (150 mL) for 24 h (3 times) to yield the crude extract (5.0 g). *L. stoechas* another sample (80 g) was boiled in water (150 mL) then filtered (whatman filter paper) and liquid part was extracted with ethyl acetate (100 mL × 3) with a separatory funnel to yield the ethyl acetate extract (2.0 g).

LC/TOF/MS Analysis

The quantitative analysis was carried out by LC-

TOF/MS, Agilent 62100 with ZORBAX SB-C18, 4.6 × 100 mm, 3.5 μm with injection volume 10 μL. The flow rate was 0.6 mL/min at 35 °C. The mobile phase consisted of eluent A, water with 0.1% formic acid and eluent B, acetonitrile. The gradient programme was as follow: 0-1 min, 10% B; 1-20 min, 50% B; 20-23 min, 80% B; 23-25 min, 10% B; 25-30 min, 10% B. The evaluation time was 30 min. Positive mode was used for TOF analysis and gas temperature was 325 °C. Drying gas flow was 10 mL/min and fragmentor voltage was 175 V (29).

Isolation of the essential oils

Aerial parts of the 25 g plant diluted with 250 mL distilled water were subjected to hydrodistillation for 4 h, using a Clevenger-type apparatus. The oil was stored at +4 °C for analysis.

GC and GC-MS analysis

GC analyses were carried out on a Perkin-Elmer Clarus 500 Series, in divided mode, 50:1, equipped with a flame ionization detector (FID) and a mass spectrometer-equipped BPX-5 apolar capillary column (30 m × 0.25 mm, 0.25 μm i.d.). The injection temperature was fixed and FID was executed at 250 °C. The carrier gas was helium at a rate of 1.0 mL/min. The initial column oven temperature was 50 °C and was raised to 220 °C at a rate of 8 °C/minute. In the mass spectrometer, transfer line temperature was at 250 °C, ionization energy was 70 eV. The standard components were used for the majority of the essential oil constituents and Kovats retention indices (RIs) were determined for all the sample components using the Van den Dool and Kratz equation according to homolog n-alkane series retention times.

Single-dose contact effects assay

The essential oil and methanol extract of plant were diluted with acetone to 100 μL/mL and 100 mg/mL respectively. Diluted essential oil and extract were applied to the dorsal surface of thorax of *S. granarius* and *S. oryzae* at an amount of 1 μL/insect with 50 μL Hamilton syringe. Acetone was used as control at a dozen of 1 μL/insect. For each replication, 20 insects were used and each experiment was repeated for three times. The tested insects were transferred to the 60 mm diameter clean Petri dishes filled with 5 g of wheat and incubated at 27 ± 2 °C in a dark climate chamber. The number of dead insects was recorded after 24 h. A randomized block design was operated comprising treatments and blank control (30).

Determination of Total Phenolic Content

Total phenolic contents of *L. stoechas* methanol extract and ethyl acetate extract were estimated by a colorimetric assay (31). An extract solution (100 μL, 1.0 μg/mL), distilled water (4.6 mL) and Folin-Ciocalteu reagent (100 μL) were mixed in a volumetric flask. After adding Na₂CO₃ (200 μL, 2%), reaction mixture was shaken for 2 h. The absorbance was measured at 760 nm in a spectrophotometer (Hitachi U-2900). The concentration of total phenolic compounds in extract was determined as mg gallic acid equivalent using an equation obtained from the standard gallic acid graph:

$Absorbance = 0.0051 \times Total\ phenols\ [Gallic\ acid\ eqv.(\mu g)] - 0.0057$

DPPH free radical scavenging activity

The DPPH[•] assay was used to investigate for the free radical scavenging activity of extracts (32). DPPH[•] solution (0.26 mM, 1.0 mL) was added to the various concentrations of extracts (3 mL, 20-100 μg/mL). The reaction mixture was stirred for 40 sec. The absorbance was measured at 517 nm with a spectrophotometer. The DPPH[•] scavenging activity was calculated using the equation:

$$DPPH^{\bullet}\ scavenging\ effect\ (\%) = [(A_c - A_s) / A_c] \times 100$$

in which, A_c is the absorbance of the control and A_s is the absorbance of the sample (33-35).

Reducing power

Reducing power of the *L. stoechas* extracts and standards was determined with the Oyaizu method (36). Potassium ferricyanide [$K_3Fe(CN)_6$] (1.25 mL, 1%) were treated with each extract and at different concentrations (1.25-5 μg/mL) at 50 °C for 30 min and total volume was completed to 2.5 mL with buffer solution (0.2 M, pH 6.7). The reaction mixture was stirred for 20 min. Trichloroacetic acid (1.25 mL, 10%) was added to the reaction mixture and then $FeCl_3$ (0.25 mL, 0.1%) was added. The absorbance was measured at 700 nm in a spectrophotometer. High absorbance value of the reaction mixture determined high reducing capability.

ABTS^{•+} scavenging activity

ABTS^{•+} scavenging activity of extracts, and standards

(BHT, BHA, Trolox) were determined according to the literature (37). This method is based on the capability of destroying ABTS radical cations by antioxidants at 734 nm absorption in comparison to BHA, BHT, and Trolox. Initially, ABTS^{•+} was prepared by the reaction of ABTS (2 mM) with potassium persulfate (2.45 mM) then it was stored for 6 h in dark at room temperature. Subsequently, ABTS^{•+} solution (1.0 mL) reacted with each sample solution (3.0 mL) at various concentrations (2.5-40 μg/mL). The inhibition was calculated for each concentration comparative to a blank absorbance. The decolorization rate was calculated as absorbance (734 nm) of reduction percent. The results were calculated as IC₅₀. The capability of ABTS^{•+} was calculated by the equation: $ABTS^{\bullet+}\ scavenging\ effect\ (\%) = [(A_c - A_s) / A_c] \times 100$ in which, A_c is ABTS^{•+} initial concentration and A_s is ABTS^{•+} remaining concentration in the sample.

Results

Two extraction procedures were carried out for *L. stoechas*, one is direct extraction with methanol to yield the methanol extract and the other is the ethyl acetate extraction in which, the plant was boiled in water then aqueous part was extracted with ethyl acetate. The quantification of methanol extract was determined by HPLC/TOF/MS analysis (Table 1).

The relationship between peak area and concentration was found to be linear from 25 to 1000 μg/L (ppb) for each compound. Linearity was evaluated by linear regression analysis of six points for each compound. Linear plot contains of three replicates per point. The correlation coefficients (R^2 values) were found as ≥ 0.99 .

Table 1. Quantitative analysis of *L. stoechas* methanol extract by HPLC/TOF/MS.

Composition (mg/1.0 kg plant)	<i>L. stoechas</i>	Linear regression equation	R^2	LOD μg/L	LOQ μg/L	%RSD	Recovery (%)
Gallic acid	trace	$y = 611,85x - 10726$	0.996	5.38	6.78	4.45	96.45
Gentisic acid	7.69	$y = 572.41x + 5626.5$	0.998	5.78	7.11	3.49	98.41
Catechine	trace	$y = 510.75x + 10111$	0.998	5.12	6.31	2.85	99.14
Chlorogenic acid	trace	$y = 348.27x + 3531.5$	0.997	9.04	14.01	4.25	100.76
4-hydroxybenzoic acid	67.06	$y = 402.27x - 1951.8$	0.998	6.89	9.41	3.12	97.50
Protocatechuic acid	1.49	$y = 645.26x + 3051.7$	0.999	4.42	5.54	1.25	104.20
Caffeic acid	27.85	$y = 1247.2x + 1675.9$	0.998	5.45	7.13	2.74	98.55
Vanillic acid	2.30	$y = 61.059x + 2701.5$	0.995	4.99	6.11	4.12	101.25
4-hydroxybenzaldehyde	trace	$y = 4525.7x + 109831$	0.998	6.90	9.21	2.91	95.90
rutin	trace	$y = 966.33x + 1355.3$	0.999	5.71	8.79	3.54	105.90
<i>p</i> -coumaric acid	3.25	$y = 813.51x + 3442.4$	0.999	7.33	11.11	3.14	99.85
Chicoric acid	trace	$y = 314.64x - 1519.1$	0.999	5.12	7.29	2.97	102.74
Ferulic acid	6.43	$y = 329.67x - 1895.4$	0.998	6.62	9.00	4.90	100.15
Hesperidin	trace	$y = 713.87x + 8548.6$	0.996	7.85	11.84	3.65	99.45
Apigenin-7- <i>O</i> -glucoside	trace	$y = 1125.4x + 15634$	0.998	8.75	13.44	2.21	100.86
Rosmarinic acid	80.89	$y = 786.7x - 7818.5$	0.998	6.25	7.44	3.55	102.50
Protocatechuic acid ethyl ester	trace	$y = 3379.4x + 53655$	0.998	4.60	7.89	2.66	85.75
Salicylic acid	1.20	$y = 1176.7x + 19371$	0.996	8.20	13.59	4.66	90.55
Quercetin	trace	$y = 2684x - 16457$	0.999	6.62	11.59	2.21	99.90
Naringenin	trace	$y = 3015.5x + 6664.8$	0.998	7.24	10.67	4.10	103.50
Campherol	trace	$y = 1853.9x - 2942.5$	0.999	5.52	6.29	3.35	98.75
Total phenolic (mg phenolic/kg plant)	198.16						

LOD: Limit of Detection, LOQ: Limit of Quantification, RSD: Relative Standard Deviation, R^2 : Regression Coefficient.

Table 2. Chemical composition of the essential oil of *Lavandula stoechas* L.

No.	Compounds	Retention time (min)	Retention index*	Content (%)
1	alpha-Pinene	12.714	939	1.09
2	Camphene	13.254	957	2.42
3	Dehydrosabinene	13.390	961	0.30
4	<i>p</i> -Cymene	15.657	1029	0.32
5	Limonene	15.811	1034	0.25
6	Linalool oxide	17.322	1077	0.47
7	Fenchone	18.033	1096	30.5
8	Fenchyl alcohol	18.903	1123	0.42
9	Campholenal	19.286	1134	0.40
10	Camphor	20.129	1158	48.1
11	Acetic acid, [4-(1-hydroxy-1-methylethyl)cyclohex-1-enyl] methyl ester	20.704	1174	1.87
12	<i>p</i> -Cymen-8-ol	21.304	1191	1.03
13	alpha-Terpineol	21.564	1197	0.33
14	Verbenone	22.345	1221	1.59
15	Bicyclo[3.1.1]heptane, 2,6,6-trimethyl-3-(2-propenyl)-, (1 α ,2 β ,3 α ,5 α)	22.517	1227	0.73
16	Carvone	23.437	1254	0.47
17	Borneolacetate	24.825	1294	1.79
18	Carvacrol	25.130	1302	0.51
19	Bisabolol	34.542	1607	0.35
20	tau-Muurolol	36.408	1674	5.72
21	Cadalene	36.988	1694	0.58
22	3-Keto- β -ionone	37.550	1715	0.75

*RI: Retention indices calculated against n-alkanes, % calculated from FID data.

Linear regression equations were given in Table 1. LOD and LOQ were determined using the measurements of reagent blanks spiked with low concentrations of analytic according to Eurachem Guide. The blank solution was spiked to 5 ppb standard. The LOD and LOQ were calculated from the following equations:

$$\text{LOD} = 3 \times S_0 \text{ and } \text{LOQ} = 10 \times S_0$$

The repeatability in the intra-day values (relative standard deviation, RSD %) for compounds was calculated using the corresponding peak areas of three replicate analyses at approximately 5 $\mu\text{g}/\text{kg}$ concentration level.

The trueness was tested as recovery of each compound from mixed stock standard solutions in spiked plant extracts. The recovery was assessed by means of three replicate measurements in a day. The average recovery data of the compounds were determined from the following equation and the recovery was given in Table 1.

$$\text{Recovery (100\%)} = \left(\frac{\text{Measured concentration}}{\text{Spiked concentration}} \right) \times 100$$

The concentration of compounds in the plant was obtained from either one of the corresponding calibration curves. Finally, the calculated concentrations were converted to mg/kg of plant.

As a result, rosmarinic acid (%80.89) was detected as the main constituent (Figure 1). The other major constituents were 4-hydroxybenzoic acid (%67.06), caffeic acid (%27.85), gentisic acid (%7.69), ferulic acid (%6.43), *p*-coumaric acid (%3.25), vanillic acid (%2.30), protocatechuic acid (% 1.49), and salicylic acid (%1.20). In addition, the total amount of phenolic

substance in *L. stoechas* was calculated as 198.16 (mg phenol / kg plant) (Table 1).

Twenty two chemical constituents of essential oil were presented by GC-MS analysis. Camphor (48.1%) and fenchone (30.5%) were indicated as the major components of the essential oil of the *L. stoechas* (Table 2).

Insecticidal activity of essential oil and methanol extract was investigated. The methanol extract showed 50% activity on *S. granarius*. The essential oil exhibited the higher activity on *S. oryzae* (62.9%) than that of the *S. granarius* (43.3%) (Table 3).

Table 3. Insecticidal activity of *L. stoechas* essential oil and methanol extracts on *S. granarius* and *S. oryzae*.

Plant materials	% Mortality \pm SD*	
	<i>S. granarius</i>	<i>S. oryzae</i>
Control	0.00 \pm 0.00 ^{e1}	0.00 \pm 0.00 ^c
Essential oil	43.33 \pm 0.0 ^b	62.92 \pm 0.04 ^b
methanol extract	50.00 \pm 0.2 ^b	0.56 \pm 0.42 ^c

¹ Means in column followed by a different lowercase letter are significantly different (Anova $P < 0.05$, Tukey test). *SD=Standard deviation.

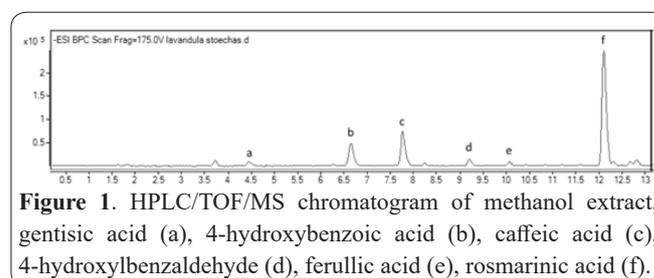
**Figure 1.** HPLC/TOF/MS chromatogram of methanol extract, gentisic acid (a), 4-hydroxybenzoic acid (b), caffeic acid (c), 4-hydroxybenzaldehyde (d), ferulic acid (e), rosmarinic acid (f).

Table 4. Antioxidant assays of *L. stoechas* extracts and standards.

Extracts	Total phenolic contents ^a	DPPH [•] scavenging activity ^b	ABTS ^{•+} Cation radical scavenging ^b	Reducing power ^c
MeOH	2.33 ± 0.016	54.60 ± 1.31	14.93 ± 0.27	0.322 ± 0.024
EtOAc	2.18 ± 0.012	28.71 ± 0.61	8.72 ± 0.21	0.699 ± 0.032
BHA		3.47 ± 0.32	3.32 ± 0.19	0.911 ± 0.027
BHT		14.78 ± 0.90	3.33 ± 0.26	0.619 ± 0.044
Trolox		4.88 ± 0.43	6.12 ± 0.18	0.467 ± 0.028

^ag gallic acid equivalent phenolic compounds/kg plant. ^bIC₅₀ values of extracts and standards (µg/ml). ^cTrolox equivalent activity (5.0 µg/ml).

Antioxidant activities of methanol and ethyl acetate extracts were carried out. Methanol extract consisted of more phenolic compounds (2.33 mg/g plant) than ethyl acetate extract (2.18 mg/g plant) whereas ethyl acetate extract exhibited better DPPH[•] scavenging activity. This might be due to the active compounds in the ethyl acetate extract. Ethyl acetate extract revealed more reducing power activity than standards, BHT and Trolox as 0.70, 0.62, 0.47 respectively at the concentration of 5.0 µg/ml. Ethyl acetate extract revealed good ABTS^{•+} scavenging activity (Table 4).

Discussion

Medical plants containing biologically active compounds are highly successful in developing new pharmaceutical components (38). Phytochemical studies from the aerial parts of *L. stoechas* have found to contain molecules such as glycoside, saponins, oleanolic acid, ursolic acid, vergatric acid, β-sitosterol, α-amylase, α-aminine acetate, luteolin, erythrodiol, flavonoids, luteolin, acacetin, vitexin, coumarin and linalool (39,40)

Caffeic acid existed in methanol extract in a significant amount (27.9 mg/1.0 kg plant) (Table 1). The caffeic acid derivatives reveal anticancer, anti-influenza activities (41). They act a considerable function in improving ionized-induced oxidative damage to red blood cells in rats, cisplatin-induced oxidative damage to the rat liver, and also regulating superoxide dismutase stress enzyme activities in diabetic rat retinas and inducing apoptosis in human multiple myeloma cells (42). Caffeic acid revealed the considerable antioxidative effect (43).

Rosmarinic acid had a number of interesting and important biological effects such as antioxidant, anti-inflammatory, anti-HIV, antimicrobial activity (44). Rosmarinic acid also includes antidepressant properties (45) and it inhibited some metabolic enzymes (46,47). It was reported that *Lavandula pedunculata subsp. lusitanica* included hydroxycinnamic acids (3-*O*-caffeoylquinic, 4-*O*-caffeoylquinic, 5-*O*-caffeoylquinic, rosmarinic acids) and flavones (luteolin and apigenin) and rosmarinic acid was the major compound (48). There was coherence with present work (Table 1, Figure 1).

The essential oil of *L. stoechas* has attracted the great interest due to its biological activities. The most commonly identified chemotype (fenchone/camphor) to that reported previously is in accordance with the present study camphor (48.1%) and fenchone (30.5%) (Table 2) were the most abundant compounds in essential oils of *L. stoechas* with one exception as different relative amounts (49-51).

Recently, extracts, essential oils, and active compounds derived from the plants gained the great interest due to the exhibiting the significant insecticidal activity. (52) Insects have the ability of developing resistance to the synthetic insecticides and have side effect on human health; therefore, bio-pesticides are preferred due to their non-toxic, biodegradable, environmental friendly effects. (53) The plants consist of bioactive compounds such as flavonoids, alkaloids, terpenoids, organic acids and lipids which are biodegradable with non-residual effects on the ecosystem. (5) Hence, the essential oils and extracts obtained from *L. stoechas* could be appropriate insecticide for stored products (Table 3).

The antioxidant activity of ethanol and water extracts of *L. stoechas* was investigated using different antioxidant analyses; superoxide anion, free radical scavenging, metal chelating, and reductive potential. The both extracts of *L. stoechas* reported to show strong antioxidant activity (54), which support this work (Table 4). In consequence of revealing the good antioxidant activity of methanol and ethyl acetate extracts, *L. stoechas* could be promising natural antioxidant agent for food industries.

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Interest conflict

No conflict of interest associated with this work.

References

- Aksit H, Çelik SM, Sen Ö, Erenler R, Demirtas I, Telci I, et al. Complete isolation and characterization of polar portion of *Mentha dumetorum* water extract. *Rec Nat Prod* 2014; 8:277-280.
- Erenler R, Sen O, Yildiz I, Aydin A. Antiproliferative activities of chemical constituents isolated from *Thymus praecox subsp grossheimii* (Ronniger) Jalas. *Rec Nat Prod* 2016; 10:766-770.
- Karan T, Erenler R. Screening of norharmane from seven cyanobacteria by high-performance liquid chromatography. *Pharmacogn Mag* 2017; 13:723-725.
- Karan T, Erenler R. Effect of Salt and pH Stress of Bioactive Metabolite Production in *Geitlerinema carotinosum*. *Int J Sec Metabol* 2017; 4:16-19.
- Demirtas I, Erenler R, Elmastas M, Goktasoglu A. Studies on the antioxidant potential of flavones of *Allium vineale* isolated from its water-soluble fraction. *Food Chem* 2013; 136:34-40.
- Elmastas M, Erenler R, Isnac B, Aksit H, Sen O, Genc N, et al. Isolation and identification of a new neo-clerodane diterpenoid from

Teucrium chamaedrys L. Nat Prod Res 2016; 30:299-304.

7. Elmastaş M, Telci İ, Akşit H, Erenler R. Comparison of total phenolic contents and antioxidant capacities in mint genotypes used as spices/Baharat olarak kullanılan nane genotiplerinin toplam fenolik içerikleri ve antioksidan kapasitelerinin karşılaştırılması. Turk J Biochem 2015; 40:456-462.
8. Topçu G, Yapar G, Türkmen Z, Gören AC, Öksüz S, Schilling JK, et al. Ovarian antiproliferative activity directed isolation of triterpenoids from fruits of *Eucalyptus camaldulensis* Dehnh. Phytochem Lett 2011; 4:421-425.
9. Ertas A, Boga M, Kizil M, Ceken B, Goren AC, Hasimi N, et al. Chemical profile and biological activities of *Veronica thymoides* subsp. *pseudocinerea*. Pharm Biol 2015; 53:334-339.
10. Davis PH. Flora of Turkey and the East Aegean Islands. Edinburgh: University Press, 1982.
11. Zuzarte M, Goncalves MJ, Cavaleiro C, Cruz MT, Benzarti A, Marongiu B et al. Antifungal and anti-inflammatory potential of *Lavandula stoechas* and *Thymus herba-barona* essential oils. Ind Crop Prod 2013; 44:97-103.
12. Kirmizibekmez H, Demirci B, Yesilada E, Baser KHC, Demirci F. Chemical Composition and Antimicrobial Activity of the Essential Oils of *Lavandula stoechas* L. ssp *stoechas* Growing Wild in Turkey. Nat Prod Commun 2009; 4:1001-1006.
13. Videira R, Castanheira P, Graos M, Salgueiro L, Faro C, Cavaleiro C. A necrodane monoterpene from *Lavandula luisieri* essential oil as a cell-permeable inhibitor of BACE-1, the β -secretase in Alzheimer's disease. Flavour Frag J 2013; 28:380-388.
14. Kulkarni RR, Joshi SP. New 2,2-diphenylpropane and ethoxylated aromatic monoterpenes from *Lavandula gibsoni* (Lamiaceae). Nat Prod Res 2013; 27:1323-1329.
15. Kurkin VA, Lamrini M. Flavonoids of *Lavandula spica* flowers. Chem Nat Compd 2007; 43:702-703.
16. Wu X, Liu J, Yu ZB, Ye YH, Zhou YW. Chemical constituents of *Lavandula augustifolia*. Acta Chim Sinica 2007; 65:1649-1653.
17. Gabrieli C, Kokkalou E. A new acetylated glucoside of luteolin and two flavone glucosides from *Lavandula stoechas* ssp *stoechas*. Pharmazie 2003; 58:426-427.
18. Topcu G, Ayril MN, Aydin A, Goren AC, Chai HB, Pezzuto JM. Triterpenoids of the roots of *Lavandula stoechas* ssp *stoechas*. Pharmazie 2001; 56:892-895.
19. Karan T, Yildiz I, Aydin A, Erenler R. Inhibition of various cancer cells proliferation of bornyl acetate and essential oil from *Inula graveolens* (Linnaeus) Desf. Rec Nat Prod 2018; 12:273-283.
20. Ertas A, Gören AC, Boga M, Demirci S, Kolak U. Chemical composition of the essential oils of three *Centaurea* species growing wild in Anatolia and their anticholinesterase activities. J Essent O Bear Pl 2014; 17:922-926.
21. Park BS, Lee SE, Choi WS, Jeong CY, Song C, Cho KY. Insecticidal and acaricidal activity of piperonaline and piperocetadecalidine derived from dried fruits of *Piper longum* L. Crop Prot 2002; 21:249-251.
22. Abay G, Karakoc OC, Tufekci AR, Koldas S, Demirtas I. Insecticidal activity of *Hypnum cupressiforme* (Bryophyta) against *Sitophilus granarius* (Coleoptera: Curculionidae). J Stored Prod Res 2012; 51:6-10.
23. Piasecka-Kwiatkowska D, Nawrot J, Zielińska-Dawidziak M, Gawlak M, Michalak M. Detection of grain infestation caused by the granary weevil (*Sitophilus granarius* L.) using zymography for α -amylase activity. J Stored Prod Res 2014; 56:43-48.
24. Koutsaviti A, Antonopoulou V, Vlassi A, Antonatos S, Michaelakis A, Papachristos DP, et al. Chemical composition and fumigant activity of essential oils from six plant families against *Sitophilus oryzae* (Col: Curculionidae). J Pest Sci 2018; 91:873-886.
25. Gulcin I. Antioxidant activity of food constituents: an overview. Arch Toxicol 2012; 86:345-391.
26. Elmastas M, Ozturk L, Gokce I, Erenler R, Aboul-Enein HY. Determination of antioxidant activity of marshmallow flower (*Althaea officinalis* L.). Anal Lett 2004; 37:1859-1869.
27. Taslimi P, Gulçin İ. Antioxidant and anticholinergic properties of olivetol. J Food Biochem 2018; 42:e12516.
28. Rezai M, Bayrak Ç, Taslimi P, Gülçin İ, Menzek A. The first synthesis and antioxidant and anticholinergic activities of 1-(4,5-dihydroxybenzyl) pyrrolidin-2-one derivative bromophenols including natural products. Turk J Chem 2018; 42:808-825.
29. Erenler R, Telci I, Ulutas M, Demirtas I, Gul F, Elmastas M, et al. Chemical constituents, quantitative analysis and antioxidant activities of *Echinacea purpurea* (L.) Moench and *Echinacea pallida* (Nutt.) Nutt. J Food Biochem 2015.
30. Polatoğlu K, Karakoç ÖC, Gören N. Phytotoxic, DPPH scavenging, insecticidal activities and essential oil composition of *Achillea vermicularis*, *A. teretifolia* and proposed chemotypes of *A. biebersteinii* (Asteraceae). Ind Crop Prod 2013; 51:35-45.
31. Singleton VL, Slinkard K. Total phenol analysis: Automation and comparison with manual methods. Am J Enol Viticult 1977; 28:49-55.
32. Blois MS. Antioxidant determinations by the use of a stable free radical. Nature 1958; 181:1199-1200.
33. Elmastas M, Celik SM, Genc N, Aksit H, Erenler R, Gulcin İ. Antioxidant activity of an Anatolian herbal tea—*Origanum minutiflorum*: isolation and characterization of its secondary metabolites. Int J Food Prop 2018; 21:374-384.
34. Han H, Yılmaz H, Gülçin İ. Antioxidant Activity of Flaxseed (*Linum usitatissimum* L.) shell and Analysis of Its Polyphenol Contents by LC-MS/MS. Rec Nat Prod 2018; 12:397-402.
35. Huyut Z, Beydemir Ş, Gülçin İ. Antioxidant and antiradical properties of selected flavonoids and phenolic compounds. Biochem Res Int 2017; 2017.
36. Oyaizu M. Studies on products of browning reaction: Antioxidative activities of products of browning reaction prepared from glucosamine. Jpn J Nutr 1986; 44:307-315.
37. Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, Rice-Evans C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. Free Radical Bio Med 1999; 26:1231-1237.
38. Palombo EA. Traditional medicinal plant extracts and natural products with activity against oral bacteria: potential application in the prevention and treatment of oral diseases. Evid Based Complement Alternat Med 2011; 2011.
39. Öztürk B, Konyalıoğlu S, Kantarci G, Çetinkol D. İzmir yöresindeki yabani *Lavandula stoechas* L. subsp. *stoechas* taksonundan elde edilen uçucu yağın bileşimi, antibakteriyel, antifungal ve antioksidan kapasitesi. Anadolu Ege Tar Araş Enst Der 2005; 15.
40. Baytop T. Türkiye'de Bitkiler ile Tedavi (Gecmiste ve Bugun). Nobel Tip Kitabevleri, İstanbul 1999.
41. Erdemli HK, Akyol S, Armutcu F, Akyol O. Antiviral properties of caffeic acid phenethyl ester and its potential application. J Intercult Ethnopharmacol 2015; 4:344-7.
42. Xie YC, Huang B, Yu KX, Xu WF. Further discovery of caffeic acid derivatives as novel influenza neuraminidase inhibitors. Bioorgan Med Chem 2013; 21:7715-7723.
43. Gülçin İ. Antioxidant activity of caffeic acid (3,4-dihydroxycinnamic acid). Toxicology 2006; 217:213-220.
44. Bulgakov VP, Inyushkina YV, Fedoreyev SA. Rosmarinic acid and its derivatives: biotechnology and applications. Critical reviews in biotechnology 2012; 32:203-217.
45. Takeda H, Tsuji M, Inazu M, Egashira T, Matsumiya T. Rosmarinic acid and caffeic acid produce antidepressive-like effect

in the forced swimming test in mice. *Eur J Pharmacol* 2002; 449:261-267.

46. Topal M, Gülçin İ. Rosmarinic acid: a potent carbonic anhydrase isoenzymes inhibitor. *Turk J Chem* 2014; 38:894-902.

47. Gülçin İ, Scozzafava A, Supuran CT, Koksal Z, Turkan F, Çetinkaya S, et al. Rosmarinic acid inhibits some metabolic enzymes including glutathione S-transferase, lactoperoxidase, acetylcholinesterase, butyrylcholinesterase and carbonic anhydrase isoenzymes. *J Enzyme Inhib Med Chem*. 2016; 31:1698-1702.

48. Costa P, Gonçalves S, Valentão P, Andrade PB, Almeida C, Nogueira JM et al. Metabolic profile and biological activities of *Lavandula pedunculata subsp. lusitanica (Chaytor) Franco*: Studies on the essential oil and polar extracts. *Food chemistry* 2013; 141:2501-2506.

49. Angioni A, Barra A, Coroneo V, Dessi S, Cabras P. Chemical composition, seasonal variability, and antifungal activity of *Lavandula stoechas* L. ssp. *stoechas* essential oils from stem/leaves and flowers. *J Agr Food Chem* 2006; 54:4364-4370.

50. Benabdelkader T, Zitouni A, Guitton Y, Jullien F, Maitre D, Casabianca H, et al. Essential oils from wild populations of Algerian *Lavandula stoechas* L.: composition, chemical variability, and in vitro biological properties. *Chem Biodivers* 2011; 8:937-953.

51. Messaoud C, Chograni H, Boussaid M. Chemical composition and antioxidant activities of essential oils and methanol extracts of three wild *Lavandula* L. species. *Nat Prod Res* 2012; 26:1976-1984.

52. Edde PA. A review of the biology and control of *Rhyzopertha dominica* (F.) the lesser grain borer. *J Stored Prod Res* 2012; 48:1-18.

53. Sundararajan G, Kumuthakalavalli R. Antifeedant activity of aqueous extract of *Gnidia glauca* Gilg. and *Toddalia asiatica* Lam. on the gram pod borer, *Helicoverpa armigera* (Hbn). *J Environ Biol* 2001; 22:11-14.

54. Gülçin İ, Şat İG, Beydemir Ş, Elmastaş M, Küfrevioğlu Öİ. Comparison of antioxidant activity of clove (*Eugenia caryophyllata* Thunb) buds and lavender (*Lavandula stoechas* L.). *Food Chemistry* 2004; 87:393-400.