

OCCURANCE AND SURVIVAL OF Vibrio alginolyticusin TAMOUDA BAY (MOROCCO)

M. SABIR^{1, 2}, N. COHEN^{2*}, A. BOUKHANJER² AND M.M. ENNAJI^{1*}

1 Laboratoire de Virologie et Hygiène & Microbiologie, Faculté des Sciences et Techniques de Mohammedia-Université Hassan II, Mohammedia – Maroc.

2 Laboratoire de Microbiologie et d'hygiène des Aliments et de l'Environnement, Institut Pasteur du Maroc,

Casablanca- Maroc.

Abstract

The objectives of this study were to investigate the spatial and seasonal fluctuations of *Vibrio alginolyticus* in marine environment of the Tamouda Bay on the Mediterranean coast of Morocco and to determine the dominant factors of the environment that govern these fluctuations. The samples (sea water, plankton, shellfish and sediment) were collected fortnightly for two years from three study sites on the coast Tamouda Bay in northern Morocco. The charge of *Vibrio* alginolyticus is determined by MPN method. The physicochemical parameters including temperature of sea water, pH, salinity, turbidity and chlorophyll a concentration were determined. Analysis of variance of specific variables and several principal component analyses showed that the temperature of seawater is the major determinant of seasonal distribution of *Vibrio alginolyticus*. The results showed a positive linear correlation between *Vibrio alginolyticus* and the water temperature, pH, turbidity and chlorophyll a. Similarly, there are seasonal variations and spatial of *Vibrio alginolyticus* in marine environment of the Tamouda bay and the highest concentrations were recorded in both years of study during the warm season whereas it was minimal during the cold season. Linear positive correlation was recorded between *Vibrio alginolyticus* populations in all ecological types of samples studied.

Key words: Environmental parameters, Mediterranean sea, Morocco, Physicochemical parameters, seasonal variation, *Vibrio alginolyticus*.

Article information's

Received on June 7, 2011 Accepted on September 1, 2011

Corresponding author

Pr Ennaji Moulay Mustapha Faculté des Sciences et Techniques de Mohammedia-Université Hassan II, Mohammedia – Maroc, B.P. 146 – Mohammedia 20650 – Morocco. Tel: +212 661 748862 Fax: +212 5 23 31 53 53 E-mail: m.ennaji@yahoo.fr

* Senior authors with equal contribution

Abbreviations: APW: Alkaline Peptone Water; °C: Celsius; **Chlo. a**: chlorophyll *a*; **Fig**: figure; **G**: gram; **H**: hour; **L**: liter; **mL**: milliliter; **\mum**: micrometer; **MPN**: Probable Number Method; **PCA**: Principal Component Analysis; r_s : Spearman coefficient; **SST**: surface seawater temperature; **TCBS**: Thiosulfate Citrate Bilesalt Sucrose; **V**.: *vibrio*; %: percent.

INTRODUCTION

Vibrio (*V*.) is a genus of bacteria indigenous to the aquatic environment. Some bacterial species of the genus Vibrio now considered emerging pathogens have been implicated in foodborne infections in Humans (12), posing a public health problem. The four species most frequently isolated in clinical microbiology laboratories V. cholerae. V_{\cdot} are V. vulnificus and V. parahaemolyticus, alginolyticus (20) which is the subject of our study. Other studies in Morocco have shown variable prevalence depending on the species of Vibrio, with a frequency of isolation especially in larger crustaceans and bivalves. (9, 13) and showed that 72% of the isolates belonged to the species Vibrio alginolyticus (14).

V. alginolyticus is a halophilic *Vibrio*, normal host mussels and was isolated as fish and seafood variety. In Humans, it was isolated from pus from ear infections and conjunctivitis,

especially after swimming in seawater. Similarly, it was shown that *Vibrio alginolyticus* should be included in the list of pathogens causing skin infections, especially if patients have been in contact with seawater in warm climate regions or with marine animals (10).

In Humans, it was isolated from pus from ear infections and conjunctivitis, skin lesions patients have been in contact with seawater, especially after bathing in seawater such was exceptionally isolated from sputum mav sometimes be responsible suppuration. Similarly, V. alginolyticus can also cause soft tissue infections such as cellulitis and necrotizing infection of the tissue and opacification of the sphenoidal sinuses (27). Moreover, this genre has been shown to be responsible for gastroenteritis in humans (17, 32). V. alginolyticus is a species with wide geographic distribution in marine and estuarine waters, particularly in bathing areas (2, 5). She has been associated with several diseases of marine animals, including fish and shellfish (4, 7 21).

Reservoirs of *V. alginolyticus* are particularly copepod zooplankton, which show the importance of research to include bacterial plankton (29). Moreover, *V. alginolyticus* is considered the most frequent species living freely in water and sediment (23) and can survive in seawater even under starvation conditions while maintaining their virulence (7).

The objectives of this study are (i) assess the hygienic quality of fish products and their environment from *Vibrio alginolyticus*, (ii) to study the seasonal effects on the burden of *Vibrio alginolyticus* in marine environment and (iii) assess the relationships between physicochemical parameters and load *Vibrio alginolyticus* in different samples.

MATERIALS AND METHODS

Description of the study area

Tamouda Bay is located in the Mediterranean coast of Morocco, between Sebta at the North $(35^{\circ}54'N, 5^{\circ}17'10"W)$ and Cap Negron at the South $(35^{\circ}40'N, 5^{\circ}16'40"W)$ (Fig. 1) where the climate is typically Mediterranean. The average annual temperature is about 18° C, while the annual rainfall average ranges between 800 and 1000 mm.

Environmental sampling

A total of six hundred forty-one (641) (236 water, 198 plankton, 67 bivalves and 130 sediment) samples were collected in three sampling sites of Tamouda Bay and analyzed for the monitoring of physicochemical parameters of seawater taken from the study area and research and counts of *V. alginolyticus* in water samples of planktons,

bivalves and sediment. Twice per month, samples were made over a period of two years (January 2007-December 2008).

One of the three coastal sites, located at the mount of Smir's river (site 2), is described as at a high risk while the two others (sites 1 and 3) are considered as at low risk. The locations of the sites were determined by using of the global positioning system (Fig. 1). For each field visit, four ecological types of samples were collected, namely, seawater, planktons, shellfishes and sediments.

Using boat, seawater samples (2L) were collected at a depth of 1m from the surface in a sterilized plastic bottles. Planktons were collected by dragging the water horizontally, at a depth of about 1m, with 200-µm-mesh plankton net (33). Shellfishes samples were purchased from local fishermen, while the sediments were collected from the surface of the coast using sterile plastic pots. After collection, samples were transported immediately to the laboratory in insulated coolers with frozen gel-packs to maintain the temperature at around 4°C. The surface seawater temperature (SST) and pH of each study site were assessed using portable instruments simultaneously to each sample collection. At the sampling sites, salinity of seawater was measured by titrimetry (6). Chlorophyll a concentration was determined from 10 or 15 ml of seawater filtered onto 0.20-µm-pore-size nitrocellulose filters (Millipore Corp., Bedford, MA, USA). The filters were frozen until extraction with acetone (25). The pigment concentration was measured with a TD700 fluorometer (Turner Designs, Sunnyvale, Calif.).

Vibrio enumeration procedure

The enumeration of V. alginolyticus our samples was carried out using the three tubes Most Probable Number Method (MPN) as described by Elliot (19) with some modifications. Briefly, 25 grams of shellfishes or sediments was inoculated into 25 mL of Alkaline Peptone Water (APW) and mixed using a stomacher for 30 seconds. Regarding seawater samples, 2L were concentrated using 0.20µm membrane filters (Millipore Corp., Bedford, MA, USA). In order to obtain a final concentrated volume corresponding to 100x the filters were washed with 20 ml APW. Planktons were used directly for V. alginolyticus enumeration. From these solutions, series of decimal dilutions were prepared in the tubes containing 9mL of APW. Each dilution was prepared in triplicate to determine the MPN. Inoculated APW was incubated at 37°C for 16-18h. The enrichment broth was then sub-cultured onto Thiosulfate Citrate Bilesalt Sucrose (TCBS) agar and incubated at 37°C for 18-24h. From each TCBS, suspected colony types yellow were picked out, streaked onto nutrient agar plus 2% NaCl to obtain pure cultures, screened for cytochrome oxidase and examined for NaCl requirement (0%, 3%, 6%, 8% and 10%). Other morphological, biochemical and cultural tests were conducted such as Gram staining, catalase reaction, and aminoacids decarboxylase reaction. Then, suspected colonies were submitted to phenotypic characterization by API 20E (bioMérieux, Marcy l'Etoile, France) employing 1% sterile saline as inoculum diluent.

Statistical Analysis

In order to compare the average values, an analysis of the variance was applied. A probability of 5% was fixed to accept or reject the null hypothesis of equality. The correlation between *V. alginolyticus* abundance and physicochemical parameters and between different environmental types of samples were carried out using



Figure 1. Map showing the geographical location of three sampling sites

Table 1. Correlation between *Vibrio alginolyticus* abundance in seawater, planktons, sediments, shellfishes and seawater temperature, seawater pH, salinity, turbidity, conductivity and chlorophyll *a*, over a 24-month period as

Variables	SST(°C)	pН	Sal.	Turb.	Cond.	Chlo.A	water	planktons	sediments	shellfishes
SST(°C)	1									
pН	0,503	1								
Salinity.	0,431	-0,097	1							
Turbidity.	-0,144	0,205	-0,383	1						
Conductivity	0,280	-0,217	0,926	-0,375	1					
Chlo.A	0,396	0,276	0,333	-0,173	0,262	1				
water	0,718	0,299	0,141	-0,028	-0,024	0,022	1			
planktons	0,908	0,447	0,377	-0,284	0,248	0,402	0,778	1		
sediments	0,866	0,363	0,319	-0,126	0,254	0,436	0,703	0,909	1	
shellfishes	0,834	0,414	0,220	-0,063	0,176	0,273	0,715	0,886	0,932	1

determined by using Pearson's rank correlation coefficient (r_s) .

SST: surface seawater temperature

Values in bold are significantly different from 0 with a significance level alpha=0,05

Table 2. Correlation between *Vibrio alginolyticus* occurrence and seawater temperature, salinity, turbidity and conductivity.

	SST(°C)	Salinity	Turbidity	Conductivity
Ν	16,5141	38,6969	1,7296	74,3212
Y	19,7170	38,7945	2,5442	74,6088
two tailed test for	<0,0001	<0,001	<0,0001	<0,0001
difference	**	**	**	**

SST: surface seawater temperature

Table 3.	Comparison	of Log ₁₀	mean	loads	Vibrio	alginolyticus	recorded	during	the	warm	season	and	cold
season.													

	Seawater	Plankton	Sediment	Shellfishes
warm season	3,30011531	7,12942837	9,3423542	9,49634585
cold season	2,2084029	5,14277092	6,75424813	6,82136376
	0,07471534	0,20256714	0,04411709	0,04374248
p-value	NS	NS	*	*

Copyright © 2011 Cellular & Molecular Biology

http://www.cellmolbiol.com

Spearman coefficient (r_s) . To determine the seasonal distribution of cultivable *V. alginolyticus* and to evaluate the effect of physicochemical parameters of seawater on the distribution of this bacterium, Principal Component Analysis (PCA) was conducted for the monthly averages of the values of *V. alginolyticus* abundance and physicochemical parameters recoded in the two sampling years. All statistical analysis methods were done using specialized statistical software.

RESULTS

Environmental parameters

The highest seawater temperature was recorded in June 2008 (23 °C) and the lowest in Decembre 2008 (15°C). Significant differences between the hot season (Mai to October) and the cold season (November to April) were found (P <0.01). Although high variability in seawater pH was observed, significant differences were found in the pH values between hot and cold seasons (P < 0.0001). There were similar fluctuations in the salinity at sites 1 and 3 during the two years, but the salinity at site 2 was consistently lower and its fluctuation was more important. Salinity was lower in site 2 during cold and rain season, associated with hurricanes, brought considerable freshwater runoff from the river. Thus. significant difference in salinity between the two seasons was found (P < 0.0001). High variability was also observed in chlorophyll a concentration and turbidity in the three sampling sites with a significant difference between hot and cold seasons (P < 0.0001 for both). The highest values of turbidity were always recorded in site 2 because also the water runoff from Smir's river, especially in cold season (Fig. 2).

Temporal and spatial variations of Vibrio alginolyticus abundance

The abundance of *V. alginolyticus* in Tamouda Bay fluctuated seasonally and ranged from undetectable to 9.9 Log_{10} MPN/g with an average abundance of 5 Log_{10} MPN/g.

V. alginolyticus showed very similar temporal patterns of abundance at the three sites but its abundance at site 2 was consistently higher. Consequently, sampling site 2 showed the significantly highest prevalence and abundance among all sites investigated, with *V. alginolyticus* present in up to 71% of the collected samples and a mean density of 5.2 Log_{10} MPN/g against 3.8 Log_{10} MPN/g and 3 Log_{10} MPN/g in sites 1 and 3 respectively (Fig. 3a). *Vibrio alginolyticus* population dynamic followed similar seasonal trend during the 2 years (Fig. 3b). The maximum abundance was observed during the hot season



Figure 2. Temporal fluctuations of physicochemical parameters in the three study sites: (a) seawater temperature, (b) seawater pH, (c) salinity, (d) turbidity and (e) chlorophyll *a*

and the minimum in cold season with significant difference between the two seasons in sediment and shellfishes (P < 0.04) (Table 3). This may be explained by the important flow of visitors in Tamouda Bay during summer period. *V. alginolyticus* showed also similar temporal patterns of abundance in different ecological types of simples. Nevertheless, the abundance in seawater was consistently lower. On the other

hand, a highly significant correlation of V. alginolyticus abundance with seawater temperature was found ($r_s = 0.718$; P < 0.0001) as well as with seawater pH, $(r_s > 0.3; P < 0.0001)$. But no significant correlation was found between salinity and Vibrio alginolyticus abundance (P> 0.05). Chlorophyll *a* and pH have also shown high significant correlation with seawater temperature ($r_s = 0.40$; P<0.0001 and $r_s = 0.5$; P < 0.0001 respectively). Moreover, a linear positive correlation was recorded between Vibrio alginolyticus populations in all ecological types of samples studied (table 1).





Figure 3. Temporal and spatial fluctuations of *Vibrio alginolyticus* (a) Temporal fluctuations per site, (b) spatial seasonal variation per ecological type of simples.

Seasonal distribution of environmental parameters

Ranking environmental variables of seawater by PCA showed monthly data divided in two groups related to seawater temperature Obtained results have (Fig. 4). clearly differentiated between cold season (November to April) and the hot one (Mai to October). Seawater temperature is the main factor determining the distribution for PCA 1 (87.1%), turbidity the main factor for PCA 2 (80.5%) and salinity the main factor for PCA 3 (50.7%). Component 1 accounted for 90.8% of the variance in the data, Component 2 for 6.2%, and Component 3 for 2.3%. This distribution in PCA variables showed gradient patterns related to seawater temperature and turbidity during the year, which indirectly explains the variation of *V*. *alginolyticus* abundance.



Figure 4. (a) Principal component plot of the environmental parameters (seawater temperature, pH, salinity, turbidity and chlorophyll a concentration). (b) Temporal distribution of environmental parameters

Seasonal distribution of Vibrio alginolyticus abundance

Ranking of months according to the abundance of *Vibrio alginolyticus* in different ecological types of samples showed similar seasonal distribution than environmental parameters supported by the first principal component, which accounted for 90.9 % of the variance in the data (Fig. 5).

DISCUSSION

Globalization is a source of risks because it led to the standardization of products and promotes the dominant players in the market at the expense of the weakest.

The environmental impacts of tourism are compounded by their seasonal and spatial concentration (coast). This encourages bacterial abundance with the risk of diseases that it may impose.





Figure 5. (a) Principal component plot of *Vibrio alginolyticus* abundance in all ecological types of samples (seawater, planktons, shellfishes and sediments). (b) Temporal distribution of *Vibrio alginolyticus* abundance.

Therefore, in this study, we have tried to contribute to the clarification of the ecological relationships between environmental parameters and *V. alginolyticus* in a Mediterranean region of the Morocco.

The results showed that the temperature of seawater is positively correlated with *V*. *alginolyticus* and accounted for 0.72 in seawater, 0.91 in plankton, 0.87 in sediment and 0.83 in shellfisc. This is consistent with other studies where a significant positive correlation was observed between the temperature of the seawater and the total *Vibrio* abundance (16, 22) Furthermore it there were seasonal fluctuations in abundance with winter minimum and a peak in August, which corresponds with the highest recorded temperatures (22° C) (26, 36).

Although pH and chlorophyll are correlated with *V. alginolyticus*, they are not directly responsible for its variability. While this study showed no correlation between the charges in *V. alginolyticus* and salinity due to the lack of variation of these parameters coastal geography where the average salinity was 35 g / L, between 30 and 41 g / L. which is consistent with the results shown in the coastal waters of the Adriatic Sea (Mediterranean) (3). However calculating correlation coefficient between *Vibrio* abundance and salinity in site 2 have shown significant correlation (rs = -0.24; P<0.05) because in this sampling site there are more fluctuations of salinity because freshwater runoff from Smir's river. The high variability of turbidity in the three sampling sites showed a significant difference between warm and cold seasons (P <0.0001 for both seasons), which is consistent with other studies that reported a positive correlation between the presence of *Vibrio spp.* and turbidity (15, 37).

This study also showed a positive linear correlation between the presence of V. alginolvticus abundance in sea water, plankton, shellfish and sediment. Moreover, our results showed that during the hot season, the load V. alginolyticus in sediments and shellfish is three times higher than in seawater. This is in total agreement with other reports where it has been demonstrated that plankton organisms are a rich source of nutrients that can enhance species of Vibrio at densities higher than the surrounding water (1, 29, 35). Other reports have shown that sediment resuspension is a major source of bacteria in water (34). Similarly, our study showed the positive association of V_{\cdot} alginolyticus in shellfish, seawater and plankton that can be explained by the ability of shellfish to filter sea water and concentrate the bacteria content in their liquor (12).

In summary, our analysis showed seasonal and spatial variations of *V. alginolyticus* in marine environment Tamouda bay. The highest concentrations were recovered during the warm seasons. The temperature was the main factor influencing the concentration of *V. alginolyticus*. These results are very important for assessing risk of infection after bathing tourists especially since Tamouda Bay is becoming a major resort center on the Mediterranean coast.

Acknowledgements - We thank all members of Royal Mounted Police brigade of M 'diq city for their help in carrying out the sampling as well as the **The VibrioSea Consortium:** Centre National d'Etudes Spatiales (CNES), France, MEDIAS-France, Toulouse, France, Collecte Localisation Satellites (CLS), Toulouse, France, Institut Pasteur (IP) Paris-France; IP Maroc; IP Algerie; IP Tunisie, Institut Agronomique et Vétérinaire Hassan II de Rabat, Morocco, Istituto di Scienze del Mare- ISMAR-CNR, Venezia, Italy, Institut Français de Recherche pour L'exploitation de la Mer (IFREMER), Brest, France, Department of Biology, University of Genova, Italy and Department of Pathology, University of Verona, Italy.

REFERENCES

1. Azandegbe, A., Garnier, M., Andrieux-Loyer, F., Kerouel, R., Philippon, X. and Nicolas J.L., Occurrence and seasonality of *Vibrio* aestuarianus in sediment and Crassostrea gigas haemolymph at two oyster farms in France. *Diseases of aquatic organisms*, 2010, 91: 213-221.

2. Baffone, W., Pianetti, A., Bruscolini, F., Barbieri, E. and Citterio, B. Occurrence and expression of virulence-related properties of *Vibrio* species isolated from widely consumed seafood products. *International Journal of Food Microbiology*, 2000, 54: 9–18.

3. Baffone, W., Tarsi, R., Pane, L., Campana, R., Repetto, B., Mariottini, G.L.and Pruzzo, C., Detection of free-living and plankton-bound vibrios in coastal waters of the Adriatic Sea (Italy) and study of their pathogenicity-associated properties. *Environmental Microbiology*, 2006, 8: 1299–1305.

4. Balebona, M.C., Andreu, M.J., Bordas, M.A., Zorrilla, I., Moriñigo, M.A. and Borrego, J.J., Pathogenicity of *Vibrio alginolyticus* from cultured gilt-head sea bream (*Sparus aurata*, L.). *Applied and Environmental Microbiology*, 1998, 64: 4269–4275.

5. Barbieri, E., Falzano, L., Fiorentini, C., Pianetti, A., Baffone, W., Fabbri, A., Matarrese, P., Casiere, A., Katouli, M., Kühni, I., Möllby, R., Bruscolini, F., and Donelli, G., Occurrence, diversity, and pathogenicity of halophilic *Vibrio spp.* and non-01 *Vibrio cholerae* from estuarine waters along the Italian Adriatic coast. *Applied and Environmental Microbiology*, 1999, 65: 2748–2753.

6. Barker, W. H. Jr., and Gangarosa, E. J., Food poisoning due to *Vibrio parahaemolyticus*. *Annu. Rev. Med.* 1974, 25: 75-81.

7. Ben Kahla-Nakbia, A., Besbesa, A., Chaieba, K., Rouabhiab, M. and Bakhroufa, A., Survival of *Vibrio alginolyticus* in seawater and retention of virulence of its starved cells. *Marine Environmental Research*, 2007, 46: 469-478.

8. Ben kahla-Nakbi, A., Chaieb, K., Besbes, A., Zmantar, T. and Bakhrouf, A., Virulence and enterobacterial repetitive intergenic consensus PCR of *Vibrio alginolyticus* strains isolated from Tunisian cultured gilthead sea bream and sea bass outbreaks. *Veterinary Microbiology*, 2006, 117: 321–327.

9. Bouchriti, N., Hamouda, A., Karib, H., Oumokhtar, B. and Yaakoubi, I., Appréciation de la qualité bactériologique des huîtres Crassostrea gigas commercialisées à Rabat. *Animalis*, 2001, 2: 26-35.

10. Campanelli, A. S., Sanchez-Politta, S. and Saurat, J.H., Cutaneous ulceration after an octopus bite: Infection due to *Vibrio alginolyticus*, an emerging pathogen. *Annales de Dermatologie et de Vénéréologie*, 2008, 135 : 225-227.

11. Carli, A., Pane, L., Casareto, L., Bertone, S. and Pruzzo, C., Occurence of *Vibrio alginolyticus* in Ligurian coast rock pools (Tyrrhenian Sea, Italy) and its association with the copepod Tigriopus fulvus (Fisher 1860). *Applied and environmental microbiology*, 1993, 59: 1960-1962.

12. China, B., De Schaetzen, M.A., Daube, G., Les mollusques bivalves, des aliments dangereux? *Ann. Méd. Vét.*, 2003, 147: 413 422.

13. Cohen, N. et Karib, H., *Vibrio spp.* dans les produits de la pêche: Risques et prévention. *Les Technomlogies de Laboratoire*, 2007, 3: 4-10.

14. Cohen, N., Karib, H., Ait Saïd, J., Lemee, L., Guenole, A. &. Quilici, M.L., Prévalence des vibrions potentiellement pathogènes dans les produits de la pêche commercialisés à Casablanca (Maroc). *Revue Méd. Vét*, 2007, 158 : 562-568.

15. Courtney, S.P., Hite, M. F., and Oliver, J. D., Ecology of *Vibrio vulnificus* in Estuarine Waters of Eastern North Carolina. *Applied and Environmental Microbiology*, 2003, 69: 3526-3531.

16. Covazzi, H. A., Marco, D. B., Massimiliano, Z., Giancarlo, A., Carla, P. and Misic, C., Vibrios in association with sedimentary crustaceans in three beaches of the northern Adriatic Sea (Italy), *Marine Pollution Bulletin*, 2008, 56: 574-579.

17. Darbas, H., Boyer, G., Jean-Pierre, H. and Riviere, M., *Vibrio alginolyticus* : isolement chez trois patients. *Médecine et maladies infectieuses*, 1992, 22 : 643-647.

18. Depaola, A., Hopkins, L. H., James, T. P, Wentz, B. and McPheaarson, R. M., Incidence of *Vibrio parahaemolyticus* in U.S. Coastal Waters and Oysters. *Applied and Environmental Microbiology*, 1990, 56: 2299-2302.

19. Elliot, E.L., Kaysner, C.A. and Tamplin, M.L.. *V.cholerae, V. parahaemolyticus, V.vulnificus* and other *Vibrio* spp. In: Bacteriological Analytical Manual, Food and Drug Administration, 7th edition. AOAC International, Arlington VA 1992:111-141.

20. Fournier, J.M., et Quilici, M.L., Infections à vibrions non cholériques. *Maladies infectieuses*, 2002, 8-026-F-15, 7 p.

21. Gómez-León, J., Villamil, L., Lemos, M.L., Novoa, B. and Figueras, A., Isolation of *Vibrio alginolyticus* and *Vibrio splendidus* from aquacultured carpet shell clam (*Ruditapes decussatus*) larvae associated with mass mortalities. *Applied and Environmental Microbiology*, 2005, 71: 98–104.

22. Gonzalez-Acosta, B., Bashan, Y., Hemandez-Saavedra, N.Y., Ascencio, F. and Cruz-Agüero, G., Seasonal temperature as the major determinant for populations of culturable bacteria in the sediments of an intact mangrove in an arid region. *FEMS Microbiology Ecology*, 2006, 55: 311-321.

23. Harriague, A., Covazzi, Brino, M. D., Zampini, M., Albertelli, G., Pruzzo, C. and Misic C., Vibrios in association with sedimentary crustaceans in three beaches of the northern Adriatic Sea (Italy). *Marine Pollution Bulletin* 2008, 56: 574-579.

24. Kaneko, T., and Colwell, R. R., Ecology of *Vibrio parahaemolyticus* in Chesapeake Bay. Journal of bacteriology, 1973, 113: 24-32.

25. Kemp, P. F., B. F. Sherr, E. B. Sherr, and J. J. Cole. Handbook of methods in aquatic microbial ecology. Lewis Publishers, *Boca Raton*, 1993, *FL*.: 1496-1497.

26. Larsen, J.L., Farid, A.F. and Dalsgaard, I., Occurrence of Vibrio parahaemolyticus and Vibrio alginolyticus in marine and estuarine bathing areas in Danish coast, Zentralbl Bakteriol Mikrobiol Hyg B., 1981, 173: 338-345.

27. Lopes, C., M., Rabadao, E. M., Ventura C., Saraiva, d.C., Côrte-Real, R. and Meliço-Silvestre A. A., A Case of *Vibrio alginolyticus* Bacteremia and Probable Sphenoiditis Following a Dive in the Sea. *Clinical Infections Diseases*, 1993, 17: 299-300.

28. Maugeri, T. L., Carbone, M., Maria, T. F. and Concetta, G., Detection and differentiation of *Vibrio vulnificus* in seawater and plankton of a coastal zone of the Mediterranean Sea. *Research in Microbiology*, 2006, 157: 194-200.

29. Montanari, M.P., Pruzzo, C., Pane, L., and Colwell, R.R., Vibrios associated with plankton in a coastal zone of the Adriatic Sea (Italy). *FEMS Microbiol Ecol* 1999, 29: 241–247.

30. Oliver, J. D., Warner, R. A., and Cleland, D. R., Distribution and ecology of *Vibrio vulnificus* and other lactose-fermenting marine vibrios in coastal waters of the

southeastern United States. Appl. Environ. Microbiol. 1982, 44: 1404-1414.

31. Oliver, J. D., Warner, R. A., and Cleland, D. R., Distribution of *Vibrio vulnificus* and other lactose-fermenting vibrios in the marine environment. *Appl. Environ. Microbiol*, 1983, 45: 985-998.

32. Reina, J., Fernandez-Baca, V. and Lopez, A., Acute gastroenteritis caused by *Vibrio alginolyticus* in an immunocompetent patient. *Clinical infectious diseases*, 1995, 21: 1044-1045

33. Signoretto, C., Burlacchini, G., Lleò, M.M., Pruzzo, C., Zampini, M., Pane, L. Adhesion of Enterococcus faecalis in the nonculturable state to plankton is the main mechanism responsible for persistence of this bacterium in both lake and seawater. *Appl Environ Microbiol.* 2004, 70: 6892–6896.

34. Stephen, F. J., Characklis, G. W., and Nobel, R. T., Sediment-water exchange of *Vibrio Sp.* And indicator bacteria: implications for persistence and transport and transport in the Neuse River Estuary, North Carolina, USA. *Water Research*, 2008, 42: 941-950.

35. Turner, J.W., Brooks, G., Dana, C. and Lipp, E. K., Plankton composition and environmental factors contribute to Vibrio seasonality. *The ISME Journal*, 2009, 3: 1082–1092.

36. Yvan, P. M., Bonnefont, J. L., Variation annuelle et identification des vibrions cultivant à 37°C dans un effluent urbain, dans des moiles et dans l'eau de mer en rade de Toulon (Méditerranée, France) *Revue canadienne de microbiologie*, 1990, 36 : 47-52.

37. Zimmerman, A. M., DePaola, A., Bowers, J. C., Krantz, J. A., Nordstrom, J. L., Johnson C. N., and Grimes, D. J., Variability of Total and Pathogenic *Vibrio parahaemolyticus* Densities in Northern Gulf of Mexico Water and Oysters. *Applied and Environmental Microbiology*, 2007, 73: 7589-7596.